

FUSARIUM WILT RESISTANCE

Test accepted: March 1991

Pathogen: *Fusarium oxysporum* Schlecht f. sp. *medicaginis* (Weimer) Snyd. & Hans.

Test authors: S. Nygaard and D. K. Barnes

PLANT CULTURE

Greenhouse

Container Bench, pot, or flat deep enough to allow root development

Media Sand or soil mixture

Temp/Light 24 to 30°C; 16+ hour daylength

No. of Plants 35 to 50 per replication

No. of Reps 3 replications minimum

Other Inoculate with *Rhizobium meliloti* Dang, encourage vigorous growth

INOCULUM CULTURE

Source Infected root tissue; axenic culture

Storage Inoculated sterile soil cultures or solid PDA medium

Temperature 0°C to ambient

Storage Life Several years in soil tubes

INOCULATION PROCEDURE

Age of Plant 8 to 10 weeks old

Type of Inoc. Microconidia in suspension

Inoc. Culture Potato or nutrient broth (2 g NaNO₃, 1 g KH₂PO₄, 0.5 g MgSO₄·7H₂O, 0.5 g KCl, 0.01 g FeSO₄·7H₂O, 0.5 g yeast extract, and 15 g sucrose per 1 L distilled water); inoculate sterile broth with PDA plugs and incubate on a shaker for 4 days at about 21°C

Concentration 1.6 X 10⁶ spores per mL or a 1:20 dilution of the shake culture

Method Bare root soak

Time of Inoc. 20 to 30 min

INCUBATION

Location Transplant to field

Plant Counts Approximately 2 weeks after transplanting, count all (alive and dead) plants as the base count for initial stand

Culture Maintain vigorous growth, control insects, clip plants once or twice.

Spacing 0.15 m between plants, 1.0 m between rows.

Age at Rating 3 months after transplanting

RATING

Plants are removed from the field and the tap root sectioned for rating.

0 Resistant No discoloration in the root

1 Resistant Small dark strands in the stele

2 Susceptible Small dark-brown arcs or rings in cross section of the stele

3 Susceptible Larger dark-brown areas, arcs or rings, or partial dark-brown ring in the outer stele

4 Susceptible Entire outer stele dark brown, plant alive

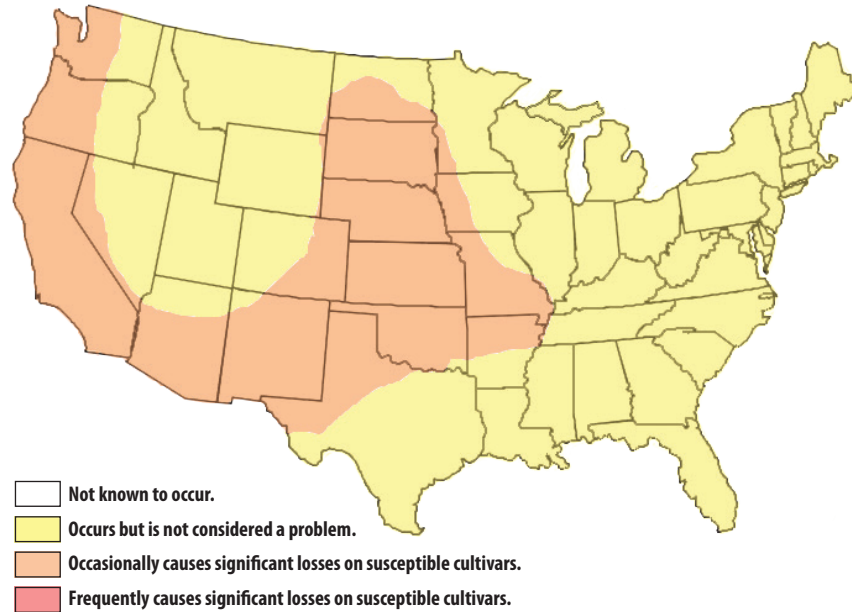
5 Susceptible Dead plant (stand loss)

CHECK CULTIVARS

	Approximate Expected Resistance (%)	Acceptable Range of Reaction (%)
Resistant		
Agate**	54	45-65
Moapa 69	76	65-85
Moderately Resistant		
Narragansett	22	15-30
Susceptible		
MNGN-1**	4	0-8

Values for resistant standards are totals of 0's and 1's.

DISTRIBUTION AND SEVERITY OF FUSARIUM WILT



Fusarium wilt, *Fusarium oxysporum* Schlecht f. sp. *Medicaginis* (Snyd. & Hans.)
(Click on the map above for a larger version.)

SOURCE OF INOCULUM AND EXPERTISE

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CULTURE OPTIONS AND RANGE OF CONDITIONS

Mixtures of *Fusarium oxysporum* and *Clavibacter michiganense* subsp. *insidiosum* inocula will not influence the severity of Fusarium wilt symptoms, but it will often produce reduced symptoms of bacterial wilt.⁽¹⁾

PLANT GROWTH OPTIONS AND RANGE OF CONDITIONS

The same procedure can be used in the greenhouse with acceptable accuracy. Plants are inoculated and then transplanted into pots, benches, or celled flats. Greenhouse evaluations can be rated 6 to 8 weeks after transplanting.

INOCULATION OPTIONS AND RANGE OF CONDITIONS

Eight to 10 week-old plants are lifted from the soil, washed in tap water, and about 50 plants for each plot are tied in a bundle. The roots are kept in tap water until all plants for all plots in a replicate are prepared. The bundled plants are immersed in the inoculum, tops trimmed to ca. 4 cm from the crown and the roots are trimmed to ca. 12 cm length. Roots are kept in moist paper towel until transplanting.

HELPFUL INFORMATION

If not planted immediately, inoculated plants may be stored at 3 to 4°C for up to a week. A tobacco planter or modified vegetable planter works well for transplanting. It can be expected that many of the highly susceptible plants will die within 5 to 6 weeks after inoculation and transplanting. Plants are undercut at 15 cm and roots are sectioned for rating. A carrot or beet lifter or sod undercutter works well for this purpose. Ratings may be expressed as an ASI or as a percentage adjusted to the long-time average of the standard check cultivar. Seed of the susceptible check (MNGN-1) can be obtained from the USDA-ARS project at the University of Minnesota.

The primary method used for evaluating resistance to Fusarium wilt has been a root soak method of inoculation followed by transplanting to the field and the scoring of root symptoms about 3 months later.^(1,2,3)

REFERENCES

1. Frosheiser, F. I., and D. K. Barnes. 1978. Field reaction of artificially inoculated alfalfa populations to the Fusarium and bacterial wilt pathogens alone and in combination. *Phytopathology* 68:943-946.
2. Frosheiser, F. I., and D. K. Barnes. 1984. In Standard tests to characterize pest resistance in alfalfa cultivars. USDA Misc. Publ. No. 1434.
3. Hijano, E. H., D. K. Barnes, and F. I. Frosheiser. 1983. Inheritance of resistance to Fusarium wilt in alfalfa. *Crop Sci.* 23:31-34.